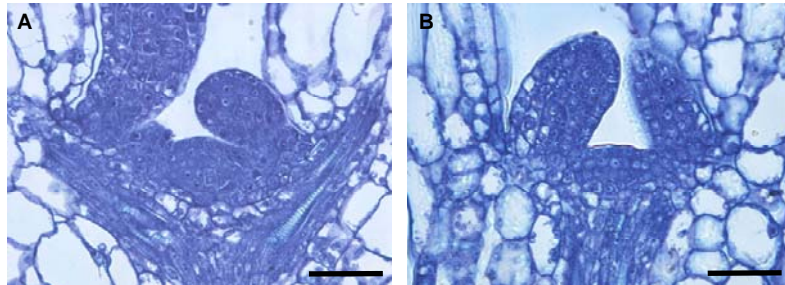


## SUPPLEMENTAL MATERIAL



**Supplemental Figure 1.** Apical meristems of En-2 and *icu4-1/icu4-1* 3-day-old seedlings show comparable size and shape. Longitudinal sections are shown from (A) En-2 and (B) *icu4-1/icu4-1* seedlings. Scale bars indicate 50  $\mu$ m.

**Supplemental Table 1.** Mutations used in this work

Mutation	Genetic background	NASC accession number	Mutagen	Origin
<i>icu4-1</i>	En-2	N400	Unknown	1
<i>icu4-2</i>	En-2	N401	Unknown	1
<i>icu4-3</i>	Col-0	N517186	T-DNA	2
<i>icu4-4</i>	Col-0	N513134	T-DNA	2
<i>athb8-3</i>	Col-0	N523733	T-DNA	2
<i>athb8-4</i>	Col-0	N565586	T-DNA	2
<i>athb8-5</i>	Col-0	N579212	T-DNA	2
<i>hst-1</i>	Ler	N3811	Diepoxybutane	3, 4
<i>crc-1</i>	Ler	N3814	EMS	5
<i>pkl-1</i>	Col*	N3840	EMS	6, 7
<i>kan1-2</i>	Ler	-	EMS	8, 9

\*As indicated in the NASC database. <sup>1</sup>Serrano-Cartagena et al. (2000). <sup>2</sup>Alonso et al. (2003). <sup>3</sup>Telfer and Poethig (1998). <sup>4</sup>Bollman et al. (2003). <sup>5</sup>Alvarez and Smyth (1999). <sup>6</sup>Ogas et al. (1997). <sup>7</sup>Ogas et al. (1999). <sup>8</sup>Eshed et al. (1999). <sup>9</sup>Kerstetter et al. (2001).

**Supplemental Table 2. Primer pairs used in this work**

Purpose	Marker/Gene	BAC	Oligonucleotide sequences (5'→3')		PCR product size (bp)
			Forward primer (label, if any)	Reverse primer	
Linkage analysis	<i>nga280</i>	F14J16	GGTGCACTTTTTATGGAGCC (6-FAM)	CTGATCTCACGGACAATAGTGC	105 (Col-0); 85 (En-2)
	<i>nga128</i>	F7A10	GGTCTGTTGATGTCGTAAGTCG (TET)	CCCTCCCTAAAGGTTTCAAGAT	162 (En-2); 181 (Col-0)
	<i>SM63</i>	F8L10	ATGCTGGACCGAAGGCTACC	CAGTCACAGAGCTCATAAACAG	178 (En-2); 154 (Col-0)
	<i>SNP375</i>	F14I3	GCACATTCACAGAAAGTTTCTG (HEX)	TACTTCATTCTGCGAAGTTAGG	201 (En-2); 219 (Col-0)
	<i>SNP17</i>	F1I21	GATGTAATGAACCCAAGTCCTG	GATTCAGGGTTTGTCTTACTG	98+171 (En-2) <sup>1</sup> ; 269 (Col-0)
	<i>F11M15</i>	F11M15	GCCGTGTTTTAATCGCCTCAG	GAGGCTTCTCAATTCTCTTTCG	839 (En-2); 832 (Col-0)
	<i>F19K6</i>	F19K6	TGGGTAAAGTGGCATTGATCAC	AATTACGAATATCAGGGCACGG	302 (En-2 and Col-0) <sup>2</sup>
Confirmation of T-DNA insertions	<i>ICU4</i>	F5F19	TATCCCTCTTGATTCCGCAAAG	LBa1 and LBb1 <sup>3</sup>	1561; 1363
	<i>ICU4</i>	F5F19	LBa1 and LBb1 <sup>3</sup>	ATGGAACCAACTTTATGGTTCAC	795; 597
	<i>ICU4</i>	F5F19	AACAATGTTCTTGCTTGGGAA	LBa1 and LBb1 <sup>3</sup>	640; 442
	<i>ICU4</i>	F5F19	LBa1 and LBb1 <sup>3</sup>	CAGAGGTGTAACGTAACAGCC	1302; 1104
	<i>ATHB-8</i>	T16I18	GCGTCGGCTCTAGACGTAGGG	LBa1 and LBb1 <sup>3</sup>	634; 436
	<i>ATHB-8</i>	T16I18	LBa1 and LBb1 <sup>3</sup>	TCTGCTTCATGTTCTCAACTGG	Not amplified
	<i>ATHB-8</i>	T16I18	AGCTTGAGAGCTTGGGGCACT	LBa1 and LBb1 <sup>3</sup>	Not amplified
	<i>ATHB-8</i>	T16I18	LBa1 and LBb1 <sup>3</sup>	GAAGCTCACACTCTCGCTCGC	682; 484
	<i>ATHB-8</i>	T16I18	CCGTCGTTGAACCTTTTAAAGACCT	LBa1 and LBb1 <sup>3</sup>	Not amplified
	<i>ATHB-8</i>	T16I18	LBa1 and LBb1 <sup>3</sup>	AGCAGGGGAATTTGGCTACCA	657; 459
RT-PCR	<i>ICU4</i>	F5F19	ATGCTAGTCCTGCAGGACTTTT	CAACCATGAGAAATAGCGATGAT	130
	<i>OTC</i>	F1B16	TGAAGGGACAAAGTTGTGTATGTT	ATCATCGCTATTTCTCATGGTTG	95

qRT-PCR	<i>YAB3</i>	F6N15	TCACGGTCACCGACAAAAGGT	GTCCTTGCTGTGAGTGTCCT	91
	<i>KAN1</i>	MQK4	GCCATGAAAGAGCAACTCCAAA	ACATGTGAAGAGCCATTTGCAG	86
	<i>KAN2</i>	F27G20	GACGACTGGATGTTTCGATATGA	AATTCTTCCGGAGAAGCACGTT	109
	<i>KNAT1</i>	F9M13	CCATTCAGGAAGCAATGGAGTT	ACTCTTCCCATCAGGATTGTTGA	101
	<i>KNAT2</i>	F24J13	CTCTTTCAGATGATGGTGCGGTT	GCGTAGTAGCTGGTCCTTCAGATC	119
	<i>KNAT6</i>	F28C11	GGGAGTTTCTGAGGATGGTGTA	TTTGAGGTCCCGGTCTTCACA	105
	<i>OTC</i>	F1B16	TGAAGGGACAAAGGTTGTGTATGTT	ATCATCGCTATTTCTCATGGTTG	95
	<i>ICU4</i>	F5F19	AGAACCGAAGATGTAGAGAGAAA	TTGACGGAAGTAGCTGTTTTTCAT	158
	<i>PNH</i>	MQD19	TCAACCAACAAGAGAGCTAATGTTT	GTGAATCCATACATCTCTTGAA	90
Genomic DNA amplification and cycle sequencing	<i>ICU4</i>	F5F19	TCAGAGAGATGCTAGTCCTGC	CAGAGGTGTAACGTAACAGCC	760 (gDNA); 386 (cDNA)
	<i>ICU4</i>	F5F19	GCTTCGTTTCGAGGTGGAATC	GTTTCTTCTGCAATGGACAAAAG	1384 (gDNA); 1058 (cDNA)
	<i>ICU4</i>	F5F19	GCGTCAGCTCAAGCAAATAGC	TCTGGCTGAGAGCTCTTAAGG	98 (gDNA and cDNA)
	<i>ICU4</i>	F5F19	TATCCCTCTTGATTCCGCAAAG	ACAAAGGACCAATTGATGAACAC	1090 (gDNA); 747 (cDNA)
	<i>ICU4</i>	F5F19	CTTCAAGGCGGGATATGTCTC	ATGGAACCAACTTTATGGTTCAC	679 (gDNA and cDNA)
	<i>ICU4</i>	F5F19	GTTTCTGAGGGAGCATAGGTC	TTCGGTTTGGGCTAGATACTTC	635 (gDNA); 389 (cDNA)
	<i>ICU4</i>	F5F19	AGAACCGAAGATGTAGAGAGAAA	TTGACGGAAGTAGCTGTTTTTCAT	158 (cDNA)
<i>In situ ICU4</i>	F5F19	TTGATGCGTATCTAGCAGCAGCAG	CGAAACGAAGCATCGATTGGAGC	435	

Oligonucleotides labeled with HEX (4, 7, 2', 4', 5', 7'-Hexachloro-6-carboxyfluorescein), TET (4, 7, 2', 7'-Tetrachloro-6-carboxyfluorescein) or 6-FAM (6-Carboxyfluorescein) phosphoramidites are indicated. Primers were designed based on the Col-0 genomic sequence available at MIPS (<http://mips.gsf.de/proj/thal/db/index.html>). <sup>1</sup>Two fragments of the indicated sizes were obtained after restriction with *HinfI* of the PCR amplification product. <sup>2</sup>The nucleotide in position 42506 of F19K6 is a T in En-2 and an A in Col-0. <sup>3</sup>PCR amplifications to confirm the presence of a T-DNA insertion were carried out using three primers, which included LBa1 (5'-TGGTTCACGTAGTGGGCCATCG-3') and LBb1 (5'-GCGTGGACCGCTTGCTGCAACT-3'). Two products of the sizes shown were obtained from each amplification.